# Impact of Lepeophtheirus salmonis infestations on migrating Atlantic salmon, Salmo salar L., smolts at eight locations in Ireland with an analysis of lice-induced marine mortality 

D Jackson ${ }^{1}$, D Cotter ${ }^{1}$, J Newell ${ }^{2}$, S McEvoy ${ }^{1}$, P O'Donohoe ${ }^{1}$, F Kane ${ }^{1}$, T McDermott ${ }^{1}$, S Kelly ${ }^{1}$ and A Drumm ${ }^{1}$<br>1 Marine Environment and Food Safety Services, Marine Institute, Galway, Ireland<br>2 School of Mathematics, Statistics and Applied Mathematics, National University of Ireland, Galway, Ireland


#### Abstract

Sea lice infestation as a source of marine mortality of outwardly migrating Atlantic salmon smolts has been investigated by treating groups of ranched salmon, prior to release, with a prophylactic sea lice treatment conferring protection from sea lice infestation. A number of studies have been carried out in Ireland using both established ranched populations and groups of hatchery reared fish imprinted for $5-8$ weeks in the sites of experimental releases. In this study, data on 352142 migrating salmon from twenty-eight releases, at eight locations along Ireland's South and West coasts covering a 9 -year period (2001 to 2009) are reviewed. Both published and new data are presented including a previously unpublished time series. The results of a meta-analysis of the combined data suggest that while sea lice-induced mortality on outwardly migrating smolts can be significant, it is a minor and irregular component of marine mortality in the stocks studied and is unlikely to be a significant factor influencing conservation status of salmon stocks.


Keywords: Atlantic salmon, Lepeophtheirus salmonis, mortality, sea lice.

[^0]
## Introduction

Across the North Atlantic region estimates of prefishery abundance of Atlantic salmon, Salmo salar L., developed by ICES indicate marked declines. The decline from the early 1970s to the mid-1990s in one-sea-winter stocks has been estimated at about $46 \%$ and in multi-sea-winter stocks at 65\% (Hutchinson \& Mills 2000). Declines in marine survival of Atlantic salmon have been recorded in Ireland (Salmon Management Task Force Report (Anon. 1996); Ó Maoilédigh et al. 2004; Jackson et al. 2011a) and elsewhere in European and North American stocks (Baum 1997; Hutchinson \& Mills 2000). The reasons for the trend towards reduced sea survival remains unclear and global warming effects (Friedland et al. 2000; Friedland, Chaput \& MacLean 2005), changes in locations or availability of prey species associated with the North Atlantic oscillation (Reid \& Planque 2000), loss of postsmolts as by-catch in pelagic fisheries, increased fishing pressure, predation, habitat changes, sea lice infestation (Finstad et al. 2007) and sea lice-induced mortality (Gargan et al. 2012) have been suggested.
The Marine Institute has undertaken a long-term study since 2001 to investigate if sea lice infestations were a significant factor in early marine mortality of Irish salmon smolts and to measure the inter-annual variation in the impacts of early sea lice infestations on sea survival. A number of aspects of this work using established ranched strains have been published (Jackson et al. 2011a,b). The goal of the present article is to analyse the data from a large number of fish (in excess of 350000 )
involving both multiple river systems and multiple releases covering almost a decade to take account of the inherent variability in salmon survival while assessing the extent of sea lice-induced mortality in Irish Atlantic salmon stocks.

## Materials and methods

## Experimental design

By treating experimental batches of tagged fish, prior to release, with a prophylactic dose of SLICE $^{\circledR}$, a commercial sea lice therapeutant, the fish can be protected from infestation with the salmon louse, Lepeophtheirus salmonis Kroyer, for up to 9 weeks (Copley et al. 2007; Jackson et al. 2011b). The active ingredient in SLICE ${ }^{\circledR}$ is emamectin benzoate. It is an animal medicine licensed for use in Ireland as a treatment for sea lice infestation in salmon. As salmon smolts are known to migrate quickly out of the bays and into the open sea, treated smolts will have moved well offshore before the protective effects of the SLICE ${ }^{\circledR}$ treatment have worn off. Studies at Burrishoole have shown that salmon smolts have moved into coastal waters within 48 h (Moore et al. 2008) and postsmolt recapture data (Shelton et al. 1997; Dadswell et al. 2010) have shown that smolts from the study area have travelled a distance of over 700 kilometres in 7 weeks and are in an area north of Scotland and west of Norway. By comparing return rates of treated fish with untreated control fish, it is possible to differentiate any additional mortality associated with sea lice infestation in the first 6-8 weeks post-migration (Jackson et al. 2011a). This methodology has been employed on a series of releases of ranched stocks from the Burrishoole river, the Bundorragha river (Delphi) and at a number of other locations (Fig. 1) on Ireland's south and west coast (Jackson et al. 2011a,b). In addition, data published by Gargan et al. (2012) using hatchery stocks transplanted into salmon and sea trout rivers and imprinted there for 5-8 weeks has been included in the analysis together with previously unpublished data from both Burrishoole and Bundorragha (Delphi).

## Tagging, tag recovery and data analysis

Experimental batches of fish were all tagged with coded wire tags. Presmolts were microtagged according to the methods of Browne (1982),


Figure 1 Locations of release points.
whereby a 1 mm long magnetized tag, etched with a specific batch code was injected into the nose cartilage of the juvenile fish. The code identifies the origin and release circumstances of any fish subsequently recaptured. All fish were anaesthetized when tagged, and the adipose fin was removed to facilitate the identification of these fish in the recovery programme. A quality control check was made on the tagged fish to ensure that the tag had been correctly magnetized. Tagging mortality and tag loss were also estimated, and subsequent analyses were based on the number of fish migrating rather than the number of fish tagged. Information on capture location and return data of the experimental groups was gathered as part of an ongoing Irish national coded wire tag recovery programme (Browne et al. 1994; Ó Maoiléidigh et al. 2004). Prior to 2007 catches from coastal commercial fisheries (drift nets, draft nets, etc.) were monitored at 15 major salmon landing ports in Ireland. These fisheries operate between May and July inclusive, and catches were scanned consistently during this period. Over $50 \%$ of the catch landed in Ireland was sampled for tags each year. The number of tagged salmon taken in these fisheries (raised data) was estimated by multiplying the actual number of tagged salmon in each area by the ratio of the total declared salmon landings in these areas to the sample size
examined. An adjustment for non-catch fishing mortality due to losses from nets and non-reporting of catches was also applied as part of this process. This methodology, as used in the compilation of returns for ICES and NASCO, ensures the avoidance of sampling bias and the comparability of data with other national and international estimates of marine survival.

## Analyses

Two-way contingency tables were used to calculate expected returns for comparison against observed returns for each pair of treatment and control batches using the chi-squared test. The resultant $P$ values were corrected using the Bonferroni procedure for multiple tests. Regression lines with $95 \%$ confidence intervals were fitted to the data set for the treated and control groups as a first step in evaluating the trends in the data. A scatterplot of percentages with a Lowess smoother was found to give a more appropriate visual representation of the data.

## Statistical analyses

The primary analysis was carried out using the generalized logistic model, and then a secondary analysis was carried out by treating the percentages as continuous (weighted) response variables.

Comparing the percentage returning without adjusting for the fact that the percentages represent considerably different denominators limits the discriminatory power of the analysis. To overcome this one needs to allow a comparison of the proportion of fish returning (i.e. a binomial response variable) between the treated and control groups to be made, while adjusting for release year and river location and for the differing number of fish migrating for each treatment/release year/location combination. A logistic regression model was fitted to model the probability of returning as a function of treatment group and release time (and their interaction) while adjusting for the association between fish released from the same location and for the differing numbers migrating from each location and year. The best model identified was one containing an interaction between release year and treatment to adjust for the fact that the positive effect of the treatment differed across release years. A generalized mixed model was fitted to the data by the Laplace approximation, and model diagnostics were carried out by
examining plots of residuals and fitted values for goodness of fit.
A linear model (i.e. an analysis of variance) was fitted where the response variable was the percentage returns (weighted by migration) with treatment, location and release date as factors. Initially, a model containing all two- and three-way interactions between the factors was fitted, and then non-significant terms were removed based on backwards elimination.

## Results

The release locations are shown in Fig. 1, and the release groups, dates of release with numbers and return rates are given in Table 1 together with $P$ values for significance. After correction using the Bonferroni adjustment, 11 of the 28 release groups or approximately $40 \%$ showed a significant difference in return rate between treated and control groups.

Of the 352142 migrating salmon, 18208 were recovered representing a sample proportion of $5.17 \%$ ( $95 \%$ confidence interval $5.1 \%$, $5.2 \%$ ). The small margin of error in the confidence interval is a consequence of the large sample size. This result suggests that, in the population of salmon represented by the sample provided, between $5.1 \%$ and $5.2 \%$ of salmon released are likely to return. The average marine mortality over the period of the study is therefore $>94 \%$, between $94.8 \%$ and $94.9 \%$.
There was a higher proportion of salmon returning in the treated group (5.6\%) compared with the controls (4.8\%). This represents a difference of approximately $0.8 \%$ between the groups favouring those having received SLICE $^{\circledR}$ (i.e. nearly $1 \%$ higher returned in the SLICE ${ }^{\circledR}$ group). An interval estimate for the difference in population proportions returning is calculated as $0.6-0.9 \%$ (Table 2). As this interval is strictly positive (i.e. does not contain zero), there is evidence that the treatment is having a positive effect in terms of a higher proportion of salmon returning albeit the improvement over the control group or absolute difference in risk is of the magnitude of approximately $1 \%$.
There is considerable variability evident in the proportions returning between the different locations as evident in the numerical summaries in Table 1. A plot of the percentage returning within each Treatment Group by Release Year with a
Table 1 Summary data on release groups of salmon including chi-squared value and $P$ value with Bonferroni correction

| Group Name | Release date | Reference | Control |  |  | Slice |  |  | Chi-squared Test |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  | Estimated Migration number | Raised Returns | Control | Estimated Migration number | Raised Returns | Slice | $P$-value |
| Delphi 01 BUR | 02/05/2001 | Jackson et al. 2011a | 6385 | 984.8 | 15.55 | 6392 | 1216.6 | 19.11 | <0.001* |
| Delphi 01 DEL | 02/05/2001 | Jackson et al. 2011a | 6395 | 892.2 | 14.11 | 6368 | 836.1 | 13.24 | 0.176 |
| Burr 01 | 03/05/2001 | Jackson et al. 2011b | 10039 | 996.6 | 9.88 | 5496 | 565.1 | 10.28 | 0.487 |
| Burr 02 | 01/05/2002 | Jackson et al. 2011b | 5989 | 542.3 | 9.10 | 5960 | 543.7 | 9.12 | 0.89 |
| Gowla 03 | 28/04/2003 | Gargan et al. 2012 | 4822 | 20.4 | 0.42 | 4955 | 225.6 | 4.55 | <0.001* |
| Invermore 03 | 29/04/2003 | Gargan et al. 2012 | 4594 | 37.7 | 0.82 | 4589 | 88.6 | 1.93 | <0.001* |
| Burr 03 | 01/05/2003 | Jackson et al. 2011b | 4587 | 373.8 | 8.15 | 4745 | 471.1 | 9.93 | 0.003 |
| Gowla 04 | 07/04/2004 | Gargan et al. 2012 | 4699 | 91.0 | 1.94 | 4655 | 164.6 | 3.54 | <0.001* |
| Invermore 04 | 08/04/2004 | Gargan et al. 2012 | 4671 | 96.2 | 2.06 | 4653 | 105.4 | 2.27 | 0.484 |
| Erriff 04 | 12/04/2004 | Gargan et al. 2012 | 4229 | 107.9 | 2.55 | 4325 | 101.8 | 2.35 | 0.551 |
| Burr 04 | 29/04/2004 | Jackson et al. 2011b | 4369 | 398.2 | 9.11 | 4437 | 403.3 | 9.07 | 0.974 |
| Erriff 05 | 04/04/2005 | Gargan et al. 2012 | 4689 | 8.4 | 0.18 | 4659 | 171.8 | 3.69 | $<0.001^{*}$ |
| Gowla 05 | 07/04/2005 | Gargan et al. 2012 | 4735 | 317.8 | 6.71 | 4583 | 306.3 | 6.68 | 0.95 |
| Invermore 05 | 08/04/2005 | Gargan et al. 2012 | 4750 | 111.2 | 2.34 | 4716 | 195.8 | 4.15 | <0.001* |
| Delphi 05 | 26/04/2005 |  | 8893 | 831.1 | 9.35 | 8471 | 1038.4 | 12.26 | <0.001* |
| Burr 05 | 28/04/2005 | Jackson et al. 2011b | 3867 | 183.2 | 4.71 | 3793 | 253.0 | 6.67 | <0.001* |
| Lee 06 | 04/04/2006 | Jackson et al. 2011a | 5131 | 10.0 | 0.19 | 5207 | 10.0 | 0.19 | 0.974 |
| Burr 06 Apr | 26/04/2006 | Jackson et al. 2011b | 4779 | 211.0 | 4.44 | 4809 | 326.0 | 6.82 | <0.001* |
| Screebe 06 | 28/04/2006 | Jackson et al. 2011a | 9618 | 121.0 | 1.26 | 10990 | 157.0 | 1.43 | 0.29 |
| Delphi 06 | 29/04/2006 |  | 8788 | 172.4 | 1.96 | 10560 | 477.9 | 4.53 | <0.001* |
| Burr 06 May | 04/05/2006 | Jackson et al. 2011b | 8000 | 334.0 | 4.21 | 3907 | 180.0 | 4.61 | 0.276 |
| Erne 06 | 04/05/2006 | Jackson et al. 2011a | 10357 | 68.0 | 0.66 | 5752 | 70.0 | 1.22 | <0.001* |
| Burr07 | 24/04/2007 | Jackson et al. 2011b | 6784 | 440 | 6.40 | 6746 | 491 | 7.29 | 0.069 |
| Delphi 07 | 26/04/2007 |  | 9719 | 567.4 | 5.84 | 9451 | 550.8 | 5.83 | 0.986 |
| Delphi 08 DEL | 28/04/2008 |  | 10811 | 183.0 | 1.69 | 16346 | 293.0 | 1.79 | 0.54 |
| Burr 08 Apr | 29/04/2008 | Jackson et al. 2011b | 6832 | 76.0 | 1.11 | 6719 | 97 | 1.44 | 0.086 |
| Burr08 May | 06/05/2008 | Jackson et al. 2011b | 3392 | 54.0 | 1.59 | 3413 | 72 | 2.11 | 0.113 |
| Burr 09 | 28/04/2009 |  | 6640 | 300.0 | 4.47 | 6881 | 267 | 3.88 | 0.064 |

[^1]Table 2 Test and CI for two proportions

| Sample | Returns | Migrated | Sample proportions |
| :---: | :---: | :---: | :---: |
| Slice | 9680 | 173578 | 0.055767 |
| Control | 8528 | 178564 | 0.047759 |
| Difference $=$ prop (Slice) - prop (Control) |  |  |  |
| Estimate for difference: 0.00800865 |  |  |  |
| 95\% CI for difference: (0.00654450, 0.00947280 ) |  |  |  |
| Test for difference $=0$ (vs. not $=0$ ): $Z=10.72$; |  |  |  |
| Fisher's exact test: $P$-Value $=0.000$ |  |  |  |

Lowess smoother superimposed is given in Fig. 2. There is a reduction in the percentage returning by year with a large reduction evident from 2001 to 2004. There is a suggestion from the smoother that the proportion returning is higher for the SLICE ${ }^{\circledR}$ group across time but that the magnitude of the difference in proportions between the groups differs across time (i.e. there appears to be a Release Date by Treatment Group interaction). To visually assess the additional effect, if any, of the Location on the proportions returning, a plot of the percentage returning by Release Year and Location panelled by Treatment Group with a Lowess smoother superimposed, is given in Fig. 3. There is evidence that the percentage returning by Year differs between locations. The highest returns were evident in the Bundorragha (Delphi) with the lowest evident in the Erriff. The Bundorragha (Delphi) and Burrishoole appear to have a similar pattern across time and between treatment groups, while the
relationship across time and between groups differ considerably for the remaining locations.

The output from the generalized logistic model identified a significant treatment effect ( $P<0.001$ ). There is evidence of an overall treatment effect favouring the treated group as the estimate is positive (0.13072). The odds of a fish returning are 1.14:1 ( $95 \%$ confidence interval $1.07,1.21$ ) in favour of the SLICE ${ }^{\circledR}$ group. The estimated probability of a treated fish returning (averaging over all years and rivers) can be calculated from the model as 0.097 compared with an estimated probability of a control fish returning (averaging over all years and rivers) of 0.086 , an absolute difference of 0.011 . This is approximately $1 \%$ or 10 fish in a thousand. Running the model for the Bundorragha (Delphi) and Burrishoole time series of data combined also gave a significant treatment effect ( $P<0.001$ ). Running the analysis for the Bundorragha (Delphi) alone gave a significant ( $P=0.001$ ) treatment effect but not for the Burrishoole time series $(P=0.49)$. As sample size is reduced the smaller numbers may be driving the lack of significance.
The outputs of the linear model (ANOVA) containing main effects only are given in Table 3. There was evidence of significant Treatment ( $P=0.034$ ), Location ( $P<0.001$ ) and Release Date ( $P<0.001$ ) effects. Two observations were identified as unusual, namely the controls in Burrishoole in 2001 where the observed percentage return ( $9.93 \%$ ) was considered low for such a control group

Figure 2 Percentage of salmon returning for each treatment group with Lowess smoother.

Scatterplot of percentage returning vs release date


## Scatterplot of percentage returning vs release date



Figure 3 Percentage of salmon returning by location and date with Lowess smoother.

Table 3 Analysis of variance for percentage, using adjusted SS for tests

| Source | df | Seq SS | Adj SS |  | Adj MS | F | P |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Location | 7 | 1968060 | 1302641 |  | 186092 | 10.19 | 0 |
| Release date | 8 | 3743239 | 3790018 |  | 473752 | 25.95 | 0 |
| Treatment | 1 | 88165 | 88165 |  | 88165 | 4.83 | 0.034 |
| Error | 39 | 711907 | 711907 |  | 18254 |  |  |
| Total | 55 | 6511371 |  |  |  |  |  |
| $S=135.107 ; \mathrm{R}-\mathrm{Sq}=89.07 \%$; $\mathrm{R}-\mathrm{Sq}(\mathrm{adj})=84.58 \%$ |  |  |  |  |  |  |  |
| Obs | Percentage | Fit |  | SE Fit |  | Residual | St Resid |
| 1 | 9.9268 | 12.329 |  | 0.7902 |  | -2.4022 | -2.2 R |
| 24 | 19.033 | 14.2887 |  | 0.7461 |  | 4.7444 | 3.13 R |

and for the treated fish originating from Bundorragha (Delphi) in 2001 where the observed percentage return of $19.03 \%$ was considered high relative to other treated fish. A plot of the adjusted mean percentages (i.e. those summarized by the model) is given in Fig. 4. As the major effect identified was release date, the ANOVA was run for the two locations with a significant time series, Burrishoole and Bundorragha (Delphi). There was evidence of a significant Release Date ( $P<0.001$ ) effect but neither Location Effect ( $P=0.116$ ) nor Treatment Group ( $P=0.156$ ) was significant (Fig. 5).

The plot of the Location Effect (Fig. 4) suggests that the highest returns were in the Bundorragha (Delphi) (adjusted mean percentage was $7.9 \%$ ) with Erriff having the lowest (adjusted mean percentage was $0.2 \%$ ). The table of the adjusted means (i.e. adjusting for migration, treatment and release date) identifies locations that have significantly different mean percentages as those that do not share a letter in common. Bundorragha (Delphi) and Burrishoole are comparable while both are significantly different (i.e. higher) than Gowla, Invermore and Erriff. There is no evidence of a
difference between Screebe, Erne, Lee, Gowla, Invermore and Erriff.

## Discussion

The data set available for analysis in this study is both large in terms of numbers of fish and comprehensive in terms of temporal and geographic coverage. The large numbers give the resulting analysis great statistical power, with the ability to detect very small differences. In designing the experimental framework (Jackson et al. 2011b), cognisance was taken of the synergistic effect of transferring smolts between rivers of different chemical composition (Saunders et al. 1983) with short-term exposure to acid waters on survival and straying. Only line bred ranched stocks reared from egg through to smolts and released within a system were used throughout the time series studies in the Delphi and Burrishoole and in the studies presented in Jackson et al. (2011b) to ensure consistency in rates of return. In contrast, Gargan et al. 2012 transferred ranched presmolts from an alkaline river

Figure 4 ANOVA: plot of adjusted mean percentages, all data.


Main effects plot for percentage Fitted means


Figure 5 ANOVA plot of adjusted mean percentages, Burrishoole and Bundorragha (Delphi) time series.
body (Lough Corrib) to distant acidic rivers. This may account for the markedly lower survival in these groups (Fig. 4), which in certain cases (e.g. Invermore and Erriff) was an order of magnitude lower than the means for the other rivers. Recent research suggests that the effects of acid water (Staurnes et al. 1996) and the interactive effects of acidification and salmon lice infestation on post-smolt survival (Finstad et al. 2007) result in reduced survival through increased predation and straying. This may limit the value of data based on stocks relocated into acid waters before release.

The temporal and geographic coverage of the data allow for a certain confidence in the results being representative. This confidence is increased by the fact that the declines in survival recorded in this study (Fig. 5) are mirrored in the reported
national marine survival trends published in the report of the Standing Scientific Committee of the National Salmon Commission (Anon 2009). Both the analysis of all data (Fig. 4) and the analysis of the time series from the Burrishoole and Bundorragha (Delphi) catchments identify 2008 as the year with the lowest marine survival in the study period and a slight recovery in marine survival in 2009. This concurs with the national trends in marine survival.
The results show a small but significant impact of sea lice infestation on marine survival of outwardly migrating salmon. This represents an absolute difference in returns of approximately $1 \%$ in favour of treated fish. This difference was significant in just under $40 \%$ of releases using the Chi-squared test with Bonferroni adjustment (Table 1). It was sensitive to sample size, being significant in the analysis
of the total data set, but not in the Burrishoole time series alone, which makes up approximately 35$40 \%$ (depending on whether you measure this in terms of number of releases or estimated migration number) of the data set (Table 1). The observed level of marine mortality attributable to sea lice infestation is very small, both in absolute terms (approximately $1 \%$ ) and as a proportion of the overall marine mortality which in this study had a mean value $>90 \%$ at all locations. At these levels, it is unlikely to influence the conservation status of stocks and is not a significant driver of marine mortality. Recent studies have been carried out in Norway with broadly similar results (Skilbrei \& Wennevik 2006).

The results also show a major fall in marine survival over the study period. This is significant in both the combined data (Fig. 4) and in each of the time series of data from Burrishoole and Bundorragha (Delphi). This fall in marine survival is mirrored in both the treated and control groups and is the main source of variance in the data ( $F=25.95$, Table 3.). When applied to the combined time series for two catchments (Fig. 5), it was the only significant source of variance. The lack of a pattern displayed for the remaining locations may be due to a combination of small sample sizes, increased levels of straying due to use of imprinted non-native stocks and a lower recovery rate for freshwater returns due to incomplete or absent upstream trapping facilities. Previous studies (Jackson et al. 2011a,b) have shown that there was no difference between the means of treated and untreated groups at one location (Burrishoole) and that a common regression of both treated and untreated groups was highly significant. That study concluded that infestation of outwardly migrating salmon with $L$. salmonis was not implicated in the observed significant decline in survival rate. The analyses carried out here on a much larger data set with two significant time series would support this conclusion. The declines in both treated and control groups follow similar trends (Fig. 2) and when separated out (Fig. 3), it can be seen that both the Bundorragha (Delphi) and Burrishoole data follow similar trends over time and across treated and untreated groups.

The marine survival data in this study reflect the reported national data on marine survival rates for wild salmon in Ireland. There is a strong and significant trend in increasing marine mortality up to 2008 . There is no evidence to suggest that this
trend is influenced by sea lice infestation levels of outwardly migrating smolts as treated and control fish are equally affected. Sea lice-induced mortality is significant in just under $40 \%$ of the releases in the study. The level of sea lice-induced mortality is small as a proportion of the overall marine mortality rate, which is in the region of $90 \%$, and in absolute terms represents $1 \%$ ( 10 fish in a thousand).

## References

Anon. (1996) Making a New Beginning in Salmon Management-Report of the Salmon Management Task Force. Government Publications, Dublin, 68pp.
Anon. (2009) Report of the Standing Scientific Committee to the Department of Communications, Energy and Natural Resources - The Status of Irish Salmon Stocks Reports in 2009 and Precautionary Catch Advice for 2010. Inland Fisheries Ireland, Dublin. 58pp.

Baum E. (1997) Maine Atlantic Salmon: A National Treasure. Atlantic salmon unlimited, Maine, 224 pp . ISBN 0-9660496-9-1.
Browne J. (1982) Results from a New Method of Tagging Salmon - The Coded Wire Tag. Fishery Leaflet, Department of Fisheries and Forestry, Dublin, 114, 10pp.
Browne J., O'Maoileidigh N., McDermott T., Cullen A., Bond N. \& McEvoy B. (1994) High Seas and Homewater Exploitation of an Irish Reared Salmon Stock. ICES CM1994/ $\mathrm{M}: 10$, ICES[s.l.] 8+annexes pp.
Copley L., O'Donohoe P., Kennedy S., Tierney D., Naughton O., Kane F., Jackson D. \& McGrath D. (2007) Lice infestation pressures on farmed Atlantic salmon smolts (Salmo salar L.) in the west of Ireland following a SLICE ( $0.2 \%$ emamectin benzoate) treatment. Fish Veterinary Journal 9, 10-21.
Dadswell M.J., Spares A.D., Reader J.M. \& Stokesbury M.J.W. (2010) The North Atlantic subpolar gyre and the marine migration of Atlantic salmon Salmo salar: the "Merry-Go-Round" hypothesis. Journal of Fish Biology 77, 435-467.

Finstad B., Kroglund F., Strand R., Stefansson S.O., Bjorn P.A., Rosseland B.O., Nilsen T.O., Salbu B. (2007) Salmon lice or suboptimal water quality - reasons for reduced postsmolt survival? Aquaculture 273, 374-383.

Friedland K.D., Hansen L.P., Dunkley D.A. \& MacLean J.C. (2000) Linkage between ocean climate, post-smolt growth and survival of Atlantic salmon (Salmo salar L.) in the North Sea area. ICES Journal of Marine Science 57, 419 -429.
Friedland K.D., Chaput G. \& MacLean J. (2005) The emerging role of climate in post-smolt growth of Atlantic salmon. ICES Journal of Marine Science 62, 1334-1349.
Gargan P.G., Forde G., Hazon N., Russell D.J.F. \& Todd C.D. (2012) Evidence for sea lice-induced marine mortality of Atlantic salmon (Salmo salar) in western Ireland from experimental releases of ranched smolts treated with emamectin
benzoate. Canadian Journal of Fisheries and Aquatic Sciences 69, 343-353.
Hutchinson P. \& Mills D. (2000) Environmental and biological factors influencing survival. In: The Ocean Life of Atlantic Salmon: Environmental and Biological Factors Influencing Survival (ed. by D. Mills), pp. 7-18. Blackwell Science, Oxford.
Jackson D., Cotter D., Ó Maoileidigh N., O'Donohoe P., White J., Kane F., Kelly S., McDermott T., McEvoy S., Drumm A., Cullen A. \& Rogan G. (2011a) An evaluation of the impact of early infestation with the salmon louse Lepeoptheirus salmonis on the subsequent survival of outwardly migrating Atlantic salmon smolts. Aquaculture 320, 159-163.
Jackson D., Cotter D., Ó Maoileidigh N., O'Donohoe P., White J., Kane F., Kelly S., McDermott T., McEvoy S., Drumm A. \& Cullen A. (2011b) Impact of early infestation with the salmon louse Lepeophtheirus salmonis on the subsequent survival of outwardly migrating Atlantic salmon smolts from a number of rivers on Ireland's south and west coasts. Aquaculture 319, 37-40.
Moore A., Cotter D., Quayle V., Rogan G., Poole R., Lower N. \& Privitera L. (2008) The impact of a pesticide on the physiology and behaviour of hatchery-reared Atlantic salmon, Salmo salar, smolts during the transition from fresh water to the marine environment. Fisheries Management and Ecology 15, 385-392.
Ó Maoiléidigh N., McGinnity P., Prévost E., Potter E.C.E., Gargan P., Crozier W.W., Mills P. \& Roche W. (2004) Application of pre-fishery abundance modeling and

Bayesian hierarchical stock and recruitment analysis to the provision of precautionary catch advice for Irish salmon (Salmo salar L.) fisheries. ICES Journal of Marine Science 61, 1370-1378.
Reid P.C. \& Planque B. (2000) Long-term planktonic variations and the climate of the North Atlantic. In: The Ocean Life of Atlantic Salmon: Environmental and Biological Factors Influencing Survival (ed. by D. Mills), pp. 153-169. Blackwell Science, Oxford.
Saunders R.L., Henderson E.B., Harmon P.R., Johnston C.E. \& Eales J.G. (1983) Effects of low environmental pH on smolting of Atlantic salmon (Salmo salar). Canadian Journal of Fisheries and Aquatic Sciences 40, 1203-1211.
Shelton R.G., Turrell W.R., Macdonald A., McLaren I.S. \& Nicoll N.T. (1997) Records of post-smolt Atlantic salmon, Salmo salar L., in the Faroe-Shetland channel in June 1996. Fisheries Research 31, 159-162.
Skilbrei O.T. \& Wennevik V. (2006) Survival \& growth of sea-ranched Atlantic salmon, Salmo salar L., treated against sea lice before release. ICES Journal of Marine Science 63, 1317-1325.
Staurnes M., Hansen L., Fugelli K. \& Haraldstad O. (1996) Short-term exposure to acid water impairs osmoregulation, seawater tolerance $\&$ subsequent marine survival of smolts of Atlantic salmon (Salmo salar L.). Canadian Journal of Fisheries and Aquatic Sciences 53, 1695-1704.

Received: 27 June 2012
Revision received: 5 October 2012
Accepted: 17 October 2012


[^0]:    Re-use of this article is permitted in accordance with the Terms and Conditions set out at http://wileyonlinelibrary.com/online open\#OnlineOpen_Terms

    Correspondence D Jackson, Marine Environment and Food Safety Services, Marine Institute, Rinville, Oranmore, Co. Galway, Ireland (e-mail: dave.jackson@marine.ie)

[^1]:    *Comparisons that were still significant after a Bonferroni correction.

