Trace Metal and Chlorinated Hydrocarbon Concentrations in Shellfish from Irish Waters, 2002

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ABSTRACT

Major shellfish growing areas were sampled in accordance with the monitoring requirements of Council Directive 79/923/EEC, on the quality required of shellfish waters, and Council Directive 91/492/EEC, laying down the health conditions for the production and placing on the market of live bivalve molluscs. Data for physicochemical parameters in water and trace metal levels and chlorinated hydrocarbon concentrations in shellfish are presented.

In 2002, a total of 24 samples from 22 different shellfish sites were analysed for chlorinated hydrocarbons and trace metals, including nickel and silver. The median concentration of mercury in shellfish sampled in 2002 was <0.03 mg kg⁻¹ wet weight, which is well within the European maximum limit of 0.50 mg kg⁻¹ wet weight for mercury in bivalve molluscs. The levels of lead and cadmium detected were low, with means of 0.16 and 0.33mg kg⁻¹ wet weight and maxima of 0.34 and 0.66 mg kg⁻¹ wet weight respectively, also within the respective European maximum levels of 1.50 and 1 mg kg⁻¹ wet weight. There are no internationally agreed standards or guidelines available for the remaining trace metals and chlorinated hydrocarbons in shellfish. Therefore, these results were compared with the strictest standard or guidance values for shellfish, which are applied by contracting countries to the OSPAR Convention, and were found to be well below the strictest values listed. This is evidence of the clean, unpolluted nature of Irish shellfish and shellfish producing waters.

As in previous years, the water quality from shellfish growing areas was good and conformed to the requirements of the Directive. Petroleum hydrocarbons were not visible in any of the shellfish waters or as deposits on the shellfish. This survey confirms previous studies (Glynn *et al.*, 2003a, 2003b; McGovern *et al.*, 2001; Bloxham *et al.*, 1998; Smyth *et al.*, 1997 and Nixon *et al.*, 1995, 1994, and 1991), which show that contamination from trace metals and chlorinated hydrocarbons is low in Irish shellfish aquaculture.

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Introduction

The determination of water quality, trace metal and chlorinated hydrocarbon concentrations in shellfish from Irish waters is carried out by the Marine Institute in part fulfillment of the monitoring requirements of various EU legislation, including:

- EU Council Directive 79/923/EEC on the quality required of shellfish growing waters, as implemented in Ireland by Statutory Instrument (SI) No. 200 of 1994,
- EU Directive 91/492/EEC laying down the health conditions for the production and placing on the market of live bivalve molluscs,
- Commission Regulation 466/2001/EC (as amended by Regulation 221/2002/EC). This information also contributes to the Joint Assessment and Monitoring Programme (JAMP) as required by the 1994 OSPAR convention.

Trace metals exist naturally in the environment and many including chromium, cobalt, copper, iron, manganese, molybdenum, vanadium, strontium and zinc are essential elements for living organisms. However, some trace metals such as mercury, lead and cadmium are not required for metabolic activity and can be toxic at quite low concentrations.

Although mercury, lead and cadmium occur naturally in the earth's crust, they can also be introduced into the aquatic environment from anthropogenic activities such as mining, industry and agriculture. Once in the aquatic environment these metals can be bioaccumulated in fish tissues. Due to physiological differences, certain species will concentrate mercury more readily than others (Clark *et al.*, 2001).

Polychlorinated biphenyls (PCBs) and organochlorine pesticides (OCPs) are man-made compounds that are ubiquitous air and water-borne contaminants. These are persistent pollutants with a tendency to bioaccumulate in fish tissue and biomagnify through the food chain (Clark *et al.*, 2001).

Council Directive 79/923/EEC requires that Member States designate shellfish growing areas. Monitoring of a range of physicochemical parameters in designated shellfish waters is undertaken to ensure that the quality of the edible species is maintained or enhanced. Directive 79/923/EEC was transposed into Irish legislation through SI No. 200 of 1994 and this also sets out designated shellfish waters in Ireland.

Sampling during 1993 and 1994 was carried out bi-annually for selected sites. The results of these surveys (Nixon *et al.*, 1994, 1995) showed that the quality of designated shellfish growing waters in Ireland was appreciably higher than required by the guidelines of the Directive, and therefore the frequency of monitoring was reduced to an annual basis in 1995, as is permitted under the Directive. Previous results were published by Glynn *et al.*, 2003a, 2003b; McGovern *et al.*, 2001; Bloxham *et al.*, 1998; Smyth *et al.*, 1997 and Nixon *et al.*, 1995, 1994 and 1991. This report presents the results of 24 shellfish samples (7 *C. gigas, 5 O. edulis* and *12 M. edulis*) from 22 sites in 2002, including a number of areas not formally designated.

Analyses of mercury, cadmium, chromium, copper, lead, nickel, silver, zinc and chlorinated hydrocarbons were carried out on shellfish from all of the sites. 2002 was the first year that nickel and silver were included in the suite of determinants. Based on previous results, special attention was given to sites that may be influenced by local anthropogenic sources.

With the exception of mercury, cadmium and lead (Commission Regulation 466/2001/EC as amended by Regulation 221/2002/EC) there are currently no applicable European standards for trace metals and chlorinated hydrocarbons in fishery products. Therefore, in the absence of EU standards for other contaminants in shellfish, monitoring results have been compared to strictest guidance or standard values available in other OSPAR Convention contracting countries.

Other reports on residue and contaminants monitoring in farmed and wild finfish, as well as other food safety and environmental monitoring reports are available from the Marine Institute (www.marine.ie/chem).

MATERIALS AND METHODS

Sample collection and preservation

Shellfish samples were collected from August to November 2002, with the exception of Ballysadare Bay sample, which was collected in March 2003. Figure 1 shows the areas sampled in 2002. Detailed information on the locations, date sampled, species sampled and cultivation methods etc. are shown in Appendix 1.

Where possible, approximate to each site where shellfish were sampled, temperature, salinity and pH measurements were taken *in situ*, typically at 1m depth using a Hydrolab® multiparameter probe (Minisonde®). At each location the water surface was visually examined for evidence of hydrocarbon contamination. Collected water samples were returned to the laboratory for the determination of suspended solids. One litre of water was filtered through a 0.45 µm membrane, which was washed with purified water and dried at 105°C. Results of the physico-chemical measurements are shown in Appendix 1.

Samples of the main shellfish species produced in each of the growing areas were collected; mussels consisting of 50 individuals and oysters of 25 individuals. Shellfish were depurated overnight in seawater collected from the growing area at the time of sampling. The lengths of individual shellfish were recorded prior to the soft tissue being removed from the shells to be washed and drained. The percentage meat and shell weights were calculated and recorded. The pooled soft tissue was then homogenised and approximately a 1g sub-sample was taken from the homogenate and dried at 105°C to constant weight to determine the moisture content. The remainder was divided into 2 sub-samples; one portion was freezedried for 48 hours and stored for metal analysis, the other stored at < -20°C prior to mercury and chlorinated hydrocarbon analysis.

Mercury analysis

Concentrated nitric acid (4ml) was added to approximately 0.7g of wet tissue and digested in a laboratory microwave oven (CEM MARS 5). After cooling, potassium permanganate was added until the purple colour of the solution stabilised. Sufficient hydroxylamine sulphate/sodium chloride solution was added to neutralise the excess potassium permanganate and 1ml of potassium dichromate was added as a preservative. The solution was diluted to 100ml with deionised water.

Following reduction of the samples with tin (II) chloride, mercury concentrations were determined by Cold Vapour Atomic Fluorescence Spectroscopy (CV-AFS) using a PSA Merlin Analyser.

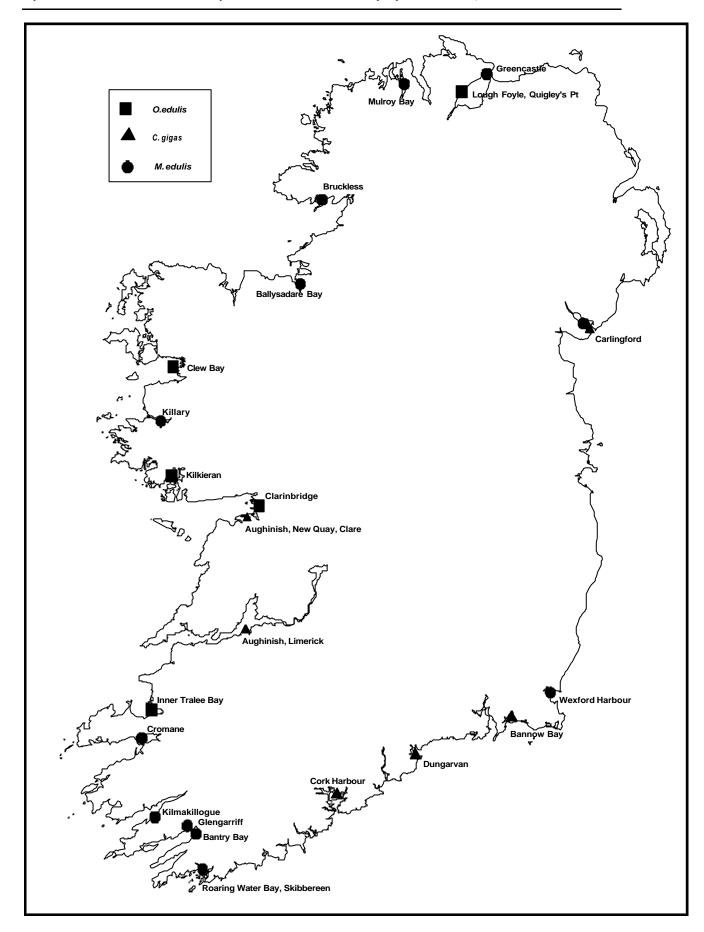


Figure 1. Location of shellfish-growing areas monitored during 2002.

Trace metal analysis (cadmium, chromium, copper, lead, zinc, silver and nickel)

Nitric acid (4ml) and hydrogen peroxide (4ml) were added to approximately 0.2g of freezedried tissue and digested in a laboratory microwave oven (CEM MARS 5). After cooling, samples were diluted to 50mls with deionised water. Lead, cadmium, chromium, copper, silver and nickel concentrations were determined using Graphite Furnace Atomic Absorption Spectrometry with Zeeman background correction (Varian SpectrAA 220Z). Zinc concentrations were determined using Flame Atomic Absorption Spectroscopy (Varian SpectrAA 20 Plus).

Chlorinated hydrocarbon analysis

Due to the lipophilic nature of PCBs and OCPs, lipid was extracted from tissue samples using the method developed by Smedes, (QUASH, 1998; QUASH, 1999). Chlorinated hydrocarbons were removed from the lipid by alumina column chromatography followed by separation of the majority of PCBs from the OCPs using silica column chromatography. Concentration levels were determined by gas chromatography with electron capture detection (GC-ECD) using a Varian CP3800 gas chromatograph fitted with two 50 metre 0.25mm i.d. fused silica capillary columns of different polarity, (HT8, J & W Scientific & CP-SIL 19CB, Chrompack).

Quality control

To ensure sufficiently high quality analytical data was produced during the 2002 shellfish monitoring programme, quality control samples, including reference materials (RMs), were analysed with each batch of samples. As the availability of appropriate marine certified reference materials (CRMs) is limited (de Boer and McGovern, 2001), reference materials supplied by QUASIMEME, (Quality Assurance of Information for Marine Environmental Monitoring), FRS Marine Laboratory, Aberdeen, were used to supplement the use of CRMs.

Although not certified, QUASIMEME provides materials of suitable matrix and analyte concentrations that have assigned values derived from intercalibrations involving many expert laboratories in this field. Obtaining a Z-score between -2 and +2 is considered satisfactory for environmental monitoring programmes.

Between 3 and 15 analyses were carried out on each RM used in this programme, the results of which are shown in Table 1. The quality assurance results obtained were considered sufficient for the purpose of the monitoring programme.

A correction factor was applied to determinants PCB 28, 31 and 52 based on recoveries for reference materials, which demonstrated consistent positive bias for these determinants. Notwithstanding, as levels are over 2 orders of magnitude less than the lowest (German) human health standard available, this bias is not considered of importance in relation to the results

Table 1: Results of the analyses of reference materials obtained during the 2002 shellfish testing.

a) Certified Reference Materials

Reference Material	Certified Values	Measured Value	No. of	Mean Z	No.
		$(Mean \pm SD)$	Analyses	Score	Z <2
Mussel Tissue CRM 278					
	mg kg ⁻¹ wet wt.	mg kg ⁻¹ wet wt.			
Cadmium	0.348	0.30 ± 0.09	12	-0.68	12
Copper	9.45	8.39 ± 0.68	10	-0.86	10
Chromium	0.78	0.60 ± 0.07	15	-1.47	12
Lead	2.00	1.77 ± 0.19	8	-0.84	8
Mercury	0.196	0.16 ± 0.01	6	-0.92	6
Zinc	83.1	79.2 ± 4.09	7	-0.35	7
Oyster Tissue SRM 156	66b				
3	mg kg ⁻¹ wet wt.	mg kg ⁻¹ wet wt.			
Cadmium	2.48	2.60 ± 0.14	9	0.36	9
Copper	71.6	71.2 ± 1.48	8	-0.04	8
Lead	0.308	0.26 ± 0.04	6	-0.75	6
Mercury	0.0371	0.025 ± 0.01	3	-0.83	3
Nickel	1.04	0.98 ± 0.10	11	-0.39	11
Silver	0.666	0.66 ± 0.03	12	-0.09	12
Zinc	1424	1559 ± 62.6	6	0.76	6

b) QUASIMEME Reference Materials

Reference Material	Assigned	Measured Value	No. of	Mean Z	No.	
	Values	$(Mean \pm SD)$	Analyses	Score	Z <2	
QOR069BT (Wet mussel tis	ssue)					
<i>PCBs</i>	mg kg ⁻¹ wet wt.	$mg kg^{-1} wet wt.$				
PCB 28	0.19	0.14 ± 0.01	4	-0.73	4	
PCB 31	0.17	0.12 ± 0.01	4	-0.72	4	
PCB 52	0.44	0.30 ± 0.03	4	-1.29	4	
PCB 101	1.89	1.68 ± 0.15	4	-0.72	4	
PCB 105	0.42	0.39 ± 0.01	4	-0.28	4	
PCB 118	1.68	1.65 ± 0.04	4	-0.10	4	
PCB 138	5.09	3.98 ± 0.27	4	-1.61	4	
PCB 153	6.60	6.08 ± 0.18	4	-0.59	4	
PCB 156	0.18	0.08 ± 0.03	4	-1.40	4	
PCB 180	0.31	0.16 ± 0.08	4	-1.68	2	
Organochlorine Pesticides						
DDD- p,p'	1.13	0.07 ± 0.13	4	-2.37	2	
DDE- p,p'	2.33	2.18 ± 0.10	4	-0.45	4	
trans-Nonachlor	0.15	0.10 ± 0.04	4	-0.69	4	
Lipid Smedes (%)	2.8	2.8 ± 0.12	4	0.08	4	

RESULTS AND DISCUSSION

Shellfish survey

The results of the biological measurements and physicochemical monitoring carried out during 2002 are given in Appendix 1. Generally, the water quality in all areas is good and conforms to the Annex of Directive 79/923/EEC.

Parameters such as pH, temperature, suspended solids and salinity are typical of results from previous monitoring. During sample collection the water surface was examined for the presence of visible petroleum hydrocarbons. No visible hydrocarbon film or deposition was evident at any of the shellfish-growing areas.

The concentrations of mercury, trace metals and chlorinated hydrocarbon contaminants analysed in the shellfish tissue are presented in Appendix 1. The level of contaminants in shellfish is a good indicator of contaminant levels present in the water column and can provide valuable information on the quality of the shellfish and the waters in which they are grown. As such, Irish shellfish monitoring data has been used for environmental assessments (Boelens *et al.*, 1999; EPA, 2000) as well as for the protection of consumers of Irish seafood products.

Commission Regulation 466/2001/EEC (as amended by Regulation 221/2002/EC) came into effect on 5th April 2002. This provides maximum levels for mercury, lead and cadmium in foodstuffs, including bivalve mussels. For mercury in fisheries products, the provisions of a previous decision (Commission Decision 93/351/EC) were transferred into this regulation. However, the maximum limit of 0.5mg kg⁻¹ wet weight is unchanged with respect to shellfish. A maximum level of 1 mg kg⁻¹ and 1.5 mg kg⁻¹ wet weight for cadmium and lead respectively in bivalve molluscs is specified in this regulation.

As there are no currently applicable European standards for contaminants in shellfish, with the exception of mercury, cadmium and lead, the levels were compared with the available standards and guidance values set for human consumption by various OSPAR contracting countries. Individual values differ between countries, but the strictest guidance and standard values are presented in Table 2.

Table 2: Synopsis of the strictest guidance and standard values applied by various OSPAR countries for contaminants in shellfish for the assessment of the possible hazards to human health (OSPAR Update 1992), updated to incorporate new European legislation on mercury, cadmium and lead.

Contaminant	Values and Units (wet weight)	Qualifier	Country
Cadmium	$1.0~{ m mg~kg}^{-1}$	Guidance	EC ¹
Copper	20 mg kg $^{-1}$	Standard	Spain ²
Lead	1.5 mg kg $^{-1}$	Guidance	EC^1
Mercury	0.5 mg kg^{-1}	Standard	EC^1
p,p' DDT and metabolites	$500~\mu g$ kg $^{-1}$	Standard	Finland
НСВ	$50~\mu g~kg^{-1}$	Guidance	Norway
α and β HCH	$50~\mu \mathrm{g~kg}^{-1}$	Guidance	Norway
Lindane	$100~\mu \mathrm{g}~\mathrm{kg}^{-1}$	Standard	Finland
PCB 28	$80~\mu \mathrm{g}~\mathrm{kg}^{-1}$	Standard	Germany
PCB 52	$80~\mu \mathrm{g}~\mathrm{kg}^{-1}$	Standard	Germany
PCB 101	$80~\mu \mathrm{g}~\mathrm{kg}^{-1}$	Standard	Germany
PCB 138	$100~\mu g$ kg $^{-1}$	Standard	Germany
PCB 153	$100~\mu g$ kg $^{-1}$	Standard	Germany
PCB 180	$80~\mu\mathrm{gkg}^{-1}$	Standard	Germany

Notes: 1. Commission Regulation 466/2001/EC as amended by Commission Regulation 221/2002/EC

Oysters are known to accumulate high levels of zinc with concentrations as high as 11,000 mg kg⁻¹ wet weight being found in the digestive glands (Clark *et al.*, 2001). The UK is the only country at present to set down a guideline value of 50 mg kg⁻¹ for Zn in food; however this excludes shellfish. The level in oysters is expected to be well in excess of 100 mg kg⁻¹ wet weight, (Anon., 1993).

2002 was the first year that nickel and silver were included in the suite of determinants. Chromium, silver and nickel contaminations in shellfish result mainly from human activities. However there are no published guidelines for acceptable concentrations of these contaminants in shellfish. Therefore, results are compared against other areas to assess for any obviously elevated results. Oysters accumulate silver to a higher concentration than mussels and this is evident from the results obtained. No specific growing area stands out as having notably elevated levels of these metals in comparison with other areas. As a larger database of results is obtained, establishment of background levels of these metals in shellfish from Irish coastal waters will enable more effective assessments in future years.

^{2.} This value does not apply to oysters for which a higher value of 60 mg kg⁻¹ has been set.

Assessment of data for individual shellfish growing areas

Data obtained during monitoring in 2002 is examined below for each sampling location and considered with respect to human consumption standards and guidance values and also to previous years monitoring data. This data is available in Fisheries Research Centre (FRC) fisheries leaflets and Marine Institute (MI) Marine Environmental Health Services publications (Glynn *et al.*, 2003a, 2003b; McGovern *et al.*, 2001; Bloxham *et al.*, 1998; Smyth *et al.*, 1997 and Nixon *et al.*, 1995, 1994 and 1991).

Aughinish, Shannon Estuary

C. gigas, were sampled from Aughinish, Shannon Estuary in 2002. Trace metal concentrations in shellfish were well within the tolerance levels given in Table 2. The level of mercury was low at <0.03 mg kg⁻¹ wet weight, well below the EU 0.5mg kg⁻¹ maximum limit (Table 2). Copper levels for C. gigas samples were found to be 46.3 mg kg⁻¹ wet weight, although this is below the Spanish standard of 60 mg kg⁻¹ set for oysters, it is still higher than 2001, where copper levels were found to be 27.0 mg kg⁻¹ wet weight. However a high value of 57.9 mg kg⁻¹ wet weight for copper was obtained in 1999, therefore there will be further investigation in the future at this site. Silver accumulates to a higher level in oysters than in mussels. The highest concentration for silver in 2002 (1.49 mg kg⁻¹) was recorded for this sample, although this was only slightly higher than the level measured in Inner Tralee Bay. Levels of PCBs and organochlorine pesticides in 2002 were comparable with previous years and were well within tolerance levels compiled by OSPAR (Table 2).

Aughinish, New Quay, Galway Bay

A sample of *C. gigas* was collected from Aughinish Bay during 2002. Trace metal concentrations were well below human consumption tolerance values as compiled by OSPAR (Table 2). Cadmium was determined at 0.56 mg kg⁻¹ wet weight, which is below the EU maximum limit (1.0 mg kg⁻¹). Copper levels were found to be 5.72 mg kg⁻¹ wet weight, well below the Spanish standard of 60 mg kg⁻¹ set for oysters, this is lower then in 2001 were copper levels were found to be 22.2 mg kg⁻¹ wet weight. However this level was more typical of previous years as from 1993-2000 levels for copper were all below 9.01 mg kg⁻¹ wet weight. The level of mercury present in the *C. gigas* sample (0.03 mg kg⁻¹ wet weight) was more than 10 times lower than the EU maximum limit for mercury in fisheries products. Analysis of the sample for organochlorine substances indicated that levels were very low with respect to human consumption standards.

Ballysadare Bay

Mussel samples, *M. edulis* were collected from Ballysadare Bay in March 2003; this was the first time that this site was sampled. However, in future a sample will be taken at this location where possible as part of our sampling programme. Trace metal levels were low and well within the strictest tolerance levels available (Table 2).

Bannow Bay

A sample of *C. gigas* was collected from Bannow Bay in 2002. Mercury was determined at <0.03 mg kg⁻¹ wet weight, well below the EU 0.5 mg kg⁻¹ maximum limit (Table 2). Other trace metal levels analysed in 2002 were well within the guideline concentrations and were similar to previous years. Chlorinated hydrocarbon levels were very low and comparable with previous values.

Bantry Bay, Inner Bay

M. edulis were tested for organics and trace metals in 2002. Trace metal concentrations in shellfish were found to be very similar to previous years and well within the tolerance levels given in Table 2. Analysis of the sample for chlorinated hydrocarbons indicated levels far below strictest tolerance levels as compiled by OSPAR.

Bruckless, McSwyne's Bay, Donegal

M. edulis, were sampled from Bruckless in 2002. Trace metal concentrations in shellfish were well within the tolerance levels given in Table 2. Cadmium levels for *M. edulis* sampled were found to be 0.40 mg kg⁻¹ wet weight, although this is below the EU maximum limit (1.0 mg kg⁻¹), it is still higher than 1999 and 2000, were cadmium levels were found to be 0.14 mg kg⁻¹ wet weight and 0.10 mg kg⁻¹ wet weight respectively. Levels of PCBs and organochlorine pesticides in 2002 were comparable with previous years and were well within tolerance levels compiled by OSPAR (Table 2).

Carlingford Lough

M. edulis and *C. gigas* (2) were sampled from Carlingford Lough. The level of mercury was low at <0.03 for all three samples. Copper levels were found to be 1.05 mg kg⁻¹ wet weight for *M. edulis* and 15.4 mg kg⁻¹ wet weight and 18.8 mg kg⁻¹ wet weight for *C. gigas*, which were well below the Spanish standard of 20 mg kg⁻¹ set for mussels and 60 mg kg⁻¹ set for oysters. Analyses for other trace metals were carried out and results were very similar to previous years monitoring. Chlorinated hydrocarbons were measured and levels were low and well within the strictest standards available for PCBs.

Clarinbridge, Galway Bay

Clarinbridge native oysters, *O. edulis* were sampled for this programme in 2002. Trace metal concentrations were well below human consumption tolerance values as compiled by OSPAR (Table 2). Cadmium was determined at 0.33 mg kg⁻¹ wet weight, which is below the EU maximum limit (1.0 mg kg⁻¹). Copper levels were found to be 10.8 mg kg⁻¹ wet weight, slightly higher than that observed for copper in 2001 (7.37 mg kg⁻¹ wet weight) but still well below the Spanish standard of 60 mg kg⁻¹ set for oysters. Levels of PCBs and organochlorine pesticides in 2002 were comparable with previous years and were well within tolerance levels compiled by OSPAR (Table 2).

Clew Bay - North

Samples of native oysters, *O. edulis*, collected in 2002 were analysed for trace metals. Trace metal levels in the shellfish were low as in previous years and mercury was determined as <0.03 mg kg⁻¹ wet weight, well within the strictest tolerance levels available (Table2). Chlorinated hydrocarbons were measured and levels were low and well within the strictest standards available.

Cork Harbour

A sample of Cork Harbour oysters, *C. gigas* was collected in 2002. Water and shellfish quality were similar to previous years and conformed to the requirements of the Directive. Copper levels were found to be 24.8 mg kg⁻¹ wet weight for *C. gigas*, well below the Spanish standard of 60 mg kg⁻¹ set for oysters. Similarly, the lead concentration of 0.12 mg kg⁻¹ wet weight was well within the EU maximum limit of 1.5 mg kg⁻¹ wet weight. Levels for other trace metal and chlorinated hydrocarbon continues to be very low.

Cromane, Castlemaine Harbour

A sample of blue mussels, *M. edulis* was collected from Cromane in 2002. Cadmium was determined at 0.25 mg kg⁻¹ wet weight, which is below the EU maximum limit (1.0 mg kg⁻¹). Copper levels were found to be 2.18 mg kg⁻¹ wet weight, well below the Spanish standard of 20 mg kg⁻¹ set for mussels. Other trace metal concentrations, including mercury continue to be low and within the tolerance levels set out in Table 2. Levels of PCBs and organochlorine pesticides in 2002 were comparable with previous years and were well within tolerance levels compiled by OSPAR (Table 2).

Dungarvan

A sample of pacific oyster, *C. gigas*, collected in 2002 was analysed for trace metals. Trace metal concentrations were comparable with previous years and were well below human consumption tolerance values as compiled by OSPAR (Table 2). Analysis of the sample for organochlorine substances indicated that levels were very low with respect to human consumption standards.

Glengarriff, Bantry Bay

M. edulis, were sampled from Glengarriff in 2002. Trace metal levels including lead (0.07 mg kg⁻¹ wet weight) were low and compared favourably with the strictest tolerance levels available (Table 2). Chlorinated hydrocarbons were measured and levels were low and well within the strictest standards available for PCBs.

Greencastle, Lough Foyle

In 2002 mussel samples, *M. edulis* were sampled from Greencastle. The levels of mercury and copper in mussels were low at <0.03 mg kg⁻¹ wet weight and 2.38 mg kg⁻¹ wet weight respectively, which were well below the well below the maximum levels given in Table 2. Analyses for other trace metals were carried out and results were very similar to previous years monitoring. Levels of PCBs and organochlorine pesticides in 2002 were comparable with previous years and were well within tolerance levels compiled by OSPAR.

Kilkieran

A sample of native oysters, *O. edulis*, collected in 2002 was analysed for trace metals. The level of mercury was low at <0.03 mg kg⁻¹ wet weight, which was well below the EU 0.5 mg kg⁻¹ wet weight maximum limit (Table 2). Copper levels were found to be 12.1 mg kg⁻¹ wet weight, which is slightly higher than in 2001 (5.07 mg kg⁻¹ wet weight) but still well below the Spanish standard of 60 mg kg⁻¹ set for oysters. Other trace metal levels in the shellfish were low as in previous years and were well within the strictest tolerance levels available (Table 2). Chlorinated hydrocarbons were measured and levels were low and well within the strictest standards available for PCBs.

Killary Harbour, Inner

M. edulis were collected at Killary Inner and analysed for both trace metals and organochlorine substances. Trace metal levels in the shellfish were low as in previous years and mercury was determined as <0.03 mg kg⁻¹ wet weight, this is well within the strictest tolerance levels available (Table 2). Chlorinated hydrocarbon analysis was carried out and results were well within the strictest standards set by OSPAR countries (Table 2).

Kilmakillogue, Kenmare River

Mussel samples, *M. edulis*, were collected from Kilmakillogue in 2002. Cadmium levels for *M. edulis* sample were found to be 0.36 mg kg⁻¹ wet weight, although this is below the EU maximum limit (1.0 mg kg⁻¹), it is still slightly higher than previous years as cadmium levels

in 2000 and 2001, were found to be 0.13 mg kg⁻¹ wet weight and 0.10 mg kg⁻¹ wet weight respectively. Other trace metal levels in the shellfish were low as in previous years and were well within the strictest tolerance levels available (Table 2). Analysis indicated organochlorine levels to be very low with respect to human consumption standards

Mulroy Bay

M. edulis were collected at Mulroy Bay and analysed for both trace metals and organochlorine substances. The levels of cadmium and lead in mussels were low at 0.24 mg kg⁻¹ wet weight and <0.06 mg kg⁻¹ wet weight respectively, which were well below the maximum levels given in Table 2. Analyses for other trace metals were carried out and results were very similar to previous years monitoring. Levels of PCBs and organochlorine pesticides in 2002 were comparable with previous years and were well within tolerance levels compiled by OSPAR.

Quigley's Point, Lough Foyle

A sample of native oysters, *O. edulis*, was sampled from Lough Foyle in 2002. The level of mercury was below the limit of quantitation (<0.03 mg kg⁻¹ wet weight) and copper levels for *O. edulis* sample was found to be 27.9 mg kg⁻¹ wet weight, well below the Spanish standard of 60 mg kg⁻¹ set for oysters. Analyses for other trace metals and chlorinated hydrocarbon were carried out and results were well within the strictest tolerance levels available (Table 2).

Roaring Water Bay

A sample of blue mussels *M. edulis*, collected in 2002 was analysed for trace metals and chlorinated hydrocarbons. Copper levels were found to be 1.34 mg kg⁻¹ wet weight for *M. edulis*, well below the Spanish standard of 20 mg kg⁻¹ set for mussels. Similarly, the lead concentration of 0.18 mg kg⁻¹ wet weight was well within the EU maximum limit of 1.5 mg kg⁻¹ wet weight. Levels for other trace metal and chlorinated hydrocarbon levels continues to be very low.

Tralee Bay, Inner

A sample of *O. edulis* was collected from Inner Tralee Bay in 2002 and analysed for trace metals and chlorinated hydrocarbons. As in previous years the mercury concentration measured in the soft tissues of *O. edulis* was very low at <0.03 mg kg⁻¹ wet weight when compared with the EU maximum limit of 0.5 mg kg⁻¹ wet weight (Table 2). The level of chromium was also low (0.22 mg kg⁻¹ wet weight), this conformed with previous monitoring and was considerably lower than the exceptional elevated value observed for 2000 sample (3.11 mg kg⁻¹ wet weight). Chlorinated hydrocarbons were measured and levels were low and well within the strictest standards available for PCBs.

Wexford Harbour

M. edulis were collected from Wexford Harbour in 2002 and were analysed for both trace metals and organochlorine substances. The level of mercury in the soft tissues of mussels was below the limit of quantitation (<0.03 mg kg⁻¹ wet weight), well below EU 0.5 mg kg⁻¹ maximum limit (Table 2). In 2002 a concentration of 0.08 mg kg⁻¹ wet weight for cadmium in mussels was detected which is well below the EU maximum limit of 1.0 mg kg⁻¹ wet weight. Similarly, the lead concentration of 0.34 mg kg⁻¹ wet weight was well within the EU maximum limit of 1.5 mg kg⁻¹ wet weight. Other trace metal levels and chlorinated hydrocarbons well within the human consumption guidelines and standards set by OSPAR countries.

CONCLUSIONS

The water quality monitored in the shellfish growing areas in 2002 was good and conformed to the guidelines of the 1979 Council Directive 79/923/EC. Nickel and silver were included in shellfish monitoring for the first time in 2002.

Based on the analyses of the 2002 samples, total mercury and trace metal concentrations in shellfish from shellfish growing areas were low, which agreed with previous studies (Glynn *et al.*, 2003a, 2003b; McGovern *et al.*, 2001; Bloxham *et al.*, 1998; Smyth *et al.*, 1997 and Nixon *et al.*, 1995, 1994 and 1991). All shellfish samples tested for mercury, cadmium and lead were well within the respective limits of 0.5 mg kg⁻¹, 1 mg kg⁻¹, and 1.5 mg kg⁻¹ wet weight, as set by European Commission Regulation 466/2001/EC, (as amended by Regulation 221/2002/EC).

Chlorinated hydrocarbon concentrations continue to be very low in Irish shellfish, which again confirmed previous studies (Glynn *et al.*, 2003; McGovern *et al.*, 2001; Bloxham *et al.*, 1998; Smyth *et al.*, 1997 and Nixon *et al.*, 1995, 1994 and 1991). All results were well within the strictest standards and guidance values of OSPAR member states.

The analytical results presented in this report are indicative of the unpolluted nature of Irish waters and fisheries products with respect to environmental contaminants.

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Appendix 1 (Page 1 of 4): Results of monitoring of shellfish-growing areas - 2002.

Sample site	Aughinish, Shannon Estuary	Aughinish, New Quay, Galway Bay	Ballysadare Bay*	Bannow Bay	Bantry Bay, Inner Bay	Bruckless, McSwyne's Bay, Donegal
M.I. Reference no.	ENV 02/067	ENV 02/061	ENV 03/025	ENV 02/083	ENV 02/064	ENV 02/071
Sampling date	29/08/02	19/08/02	05/03/03	09/10/02	27/08/02	02/10/02
Sampling time	16.30	10.45	13.30	16.12	9.00	12.30
Species sampled	C. gigas	C. gigas	M. edulis	C. gigas	M. edulis	M. edulis
Number individuals	25	25	50	25	50	35
Method of cultivation	trestle	bed	bed	trestle	rope	rope
Water Parameters						
Temperature (°C)	N/A	N/A	8.78	12.6	16.7	14.5
Salinity (psu)	N/A	N/A	N/A	10.6	32.3	26.7
pH	N/A	N/A	7.51	8.20	7.68	7.93
Suspended Solids (mg L ⁻¹)	9.20	29.4	N/A	6.70	2.90	N/A
Shellfish						
Shell length range (mm)	80.4-100	84.4-125	40.0-78.4	75.9-116	46.2-60.0	47.6-63.9
Shell mean length (mm)	89.9	97.5	59.8	95.2	54.6	58.5
Shell length std dev (mm)	5.46	8.49	7.87	9.67	3.41	4.09
Meat weight (%)	7.29	14.4	25.9	24.3	42.4	40.6
Shell weight (%)	92.7	85.6	74.1	75.7	57.6	59.4
Meat water content (%)	83.3	77.1	80.5	75.7	77.1	76.5
Metals mg kg ⁻¹ (ppm) wet						
Cadmium	0.66	0.56	0.15	0.11	0.16	0.40
Chromium	< 0.19	< 0.19	< 0.19	< 0.19	< 0.19	nd
Copper	46.3	5.72	1.27	4.56	1.09	1.26
Lead	0.19	< 0.06	0.34	0.10	0.09	< 0.06
Mercury	< 0.03	0.03	< 0.03	< 0.03	nd	nd
Zinc	466	532	16.6	112	24.6	23.8
Nickel Silver	0.19 1.49	<0.14 0.14	0.21 nd	nd 0.08	0.21 <0.03	<0.14 <0.03
		0.14	IId	0.08	<0.03	<0.03
PCB's µg kg ⁻¹ (ppb) wet w		-0.01	NT/A	0.05	.0.01	0.05
CB Congener 28 CB Congener 31	<0.01 0.02	<0.01 0.02	N/A N/A	0.05 0.06	<0.01 0.05	0.05 0.06
CB Congener 52	0.02	0.02	N/A N/A		0.03	0.06
	0.11	0.10	N/A N/A	0.15 0.23	0.23	0.13
CB Congener 101 CB Congener 105	0.13	0.10	N/A N/A	0.23	0.30	0.14
	0.02	0.01	N/A N/A	0.03	0.03	0.10
CB Congener 118 CB Congener 138	0.07	0.04	N/A N/A	<0.17	0.09	< 0.09
CB Congener 153	0.12	0.03	N/A N/A	0.11	0.28	< 0.10
CB Congener 155 CB Congener 156	< 0.19	< 0.02	N/A N/A	< 0.03	< 0.06	<0.10
CB Congener 180	< 0.02	< 0.02	N/A N/A	< 0.03	0.09	< 0.03
Organic pesticides µg kg ⁻¹		10.01	1 1/1 1	10.01	0.07	
DDD - pp'	0.03	0.03	N/A	0.18	< 0.01	0.07
DDE - pp'	0.18	0.09	N/A	0.86	0.12	0.19
DDT - op'	< 0.01	0.02	N/A	< 0.02	0.04	< 0.02
DDT - pp'	< 0.03	0.07	N/A	< 0.14	0.17	< 0.12
Dieldrin	N/A	N/A	N/A	0.56	N/A	0.33
HCB	0.01	0.01	N/A	< 0.01	0.02	0.02
alpha-HCH	0.01	0.01	N/A	0.13	0.01	0.07
beta-HCH	N/A	N/A	N/A	< 0.23	N/A	< 0.20
датта-НСН	< 0.01	0.02	N/A	0.05	< 0.02	0.04
trans-Nonachlordane	< 0.01	< 0.01	N/A	0.08	< 0.03	0.05
trans-Chlordane	< 0.02	< 0.02	N/A	< 0.02	< 0.07	0.02
cis-Chlordane	n.d	n.d	N/A	< 0.02	< 0.01	< 0.02
Endrin	n.d	0.01	N/A	< 0.07	< 0.01	< 0.06
Lipid Smedes (%)	1.27	1.32	N/A	3.48	2.10	2.17

Notes: N/A: Samples not analysed

nd: Substances were not detected above the Limit of Detection (LOD) (Appendix 2)

< value: value = Limit of Quantitation (LOQ) for the relevant determinand

^{*:} Ballysadare Bay sample collected March 2003

Appendix 1 (Page 2 of 4): Results of monitoring of shellfish-growing areas - 2002.

Sample site	Carlingford Lough	Carlingford Lough	Carlingford Lough	Clarinbridge, Galway Bay	Clew Bay - North	Cork Harbour
M.I. Reference no.	ENV 02/093	ENV 02/094	ENV 02/095	ENV 02/058	ENV 02/059	ENV 02/073
Sampling date	27/11/02	27/11/02	27/11/02	19/08/02	20/08/02	07/10/02
Sampling time	9.30	11.30	9.30	12.30	13.00	9.30
Species sampled	M. edulis	C. gigas	C. gigas	O. edulis	O. edulis	C. gigas
Number individuals	50	25	25	25	25	25
Method of cultivation	trestle	trestle	trestle	bed	bed	bed
Water Parameters						
Temperature (°C)	10.2	10.9	10.2	N/A	N/A	15.7
Salinity (psu)	29.5	29.0	29.5	N/A	N/A	31.3
pH	8.03	7.85	8.03	N/A	N/A	7.81
Suspended Solids (mg L ⁻¹)	9.4	116	9.0	10.0	33.9	10.4
Shellfish						
Shell length range (mm)	45.3-59.3	76.2-127	96.8-139	64.1-107	64.5-87.5	60.4-78.8
Shell mean length (mm)	51.6	104	112	85.9	77.9	66.3
Shell length std dev (mm)	3.63	12.7	10.9	12.8	6.36	3.77
Meat weight (%)	26.5	12.5	12.1	9.40	9.22	12.3
Shell weight (%)	73.5	87.5	87.9	90.6	90.8	87.7
Meat water content (%)	79.0	78.5	76.2	71.2	78.4	74.3
Metals mg kg-1 (ppm) wet	wt.					
Cadmium	0.18	0.37	0.57	0.33	0.57	0.29
Chromium	0.25	nd	nd	nd	< 0.19	< 0.19
Copper	1.05	15.4	18.8	10.8	3.75	24.8
Lead	0.10	0.18	0.24	0.11	< 0.06	0.12
Mercury	< 0.03	< 0.03	< 0.03	0.03	< 0.03	< 0.03
Zinc	10.9	243	351	260	343	437
Nickel Silver	0.22 <0.03	nd	nd 0.70	0.17	<0.14 0.78	nd 1.21
		0.77	0.70	0.46	0.78	1.21
PCB's µg kg ⁻¹ (ppb) wet wt		0.04	0.07	0.28	ر0.01	0.06
CB Congener 28 CB Congener 31	0.06 0.05	0.04 0.06	0.07 0.08	0.28 0.13	<0.01 0.03	0.06 0.06
CB Congener 52	0.03	0.00	0.08	0.13	0.03	0.00
CB Congener 101	0.14	0.13	0.17	0.22	0.19	0.13
CB Congener 105	0.13	< 0.05	0.10	0.03	0.10	0.11
CB Congener 118	0.13	0.06	0.10	0.06	0.02	0.47
CB Congener 138	0.46	< 0.11	0.37	0.05	0.05	0.62
CB Congener 153	1.04	<0.11	0.82	0.03	0.08	1.12
CB Congener 156	< 0.02	< 0.11	< 0.03	<0.03	< 0.04	<0.02
CB Congener 180	< 0.01	< 0.01	< 0.02	<0.03	< 0.02	< 0.02
Organic pesticides µg kg ⁻¹						
DDD - pp'	0.48	< 0.02	0.24	0.11	< 0.01	0.21
DDD - pp DDE - pp'	0.48	0.06	0.60	0.11	0.11	0.21
			0.60		0.11	
DDT - op' DDT - pp'	<0.01 0.12	<0.02 <0.14	0.04 <0.16	0.03 0.15		<0.01 <0.11
DD1 - pp Dieldrin	0.12	0.13	0.16	0.15 N/A	0.18 N/A	<0.11 0.54
HCB	0.91	< 0.01	< 0.01	0.01	0.01	< 0.01
	0.02	0.04	0.13	0.01	0.01	0.12
alpha-HCH beta-HCH	< 0.17	< 0.23	< 0.26	0.01 N/A	0.01 N/A	<0.17
датта-НСН датта-НСН	0.17	<0.23 <0.02	0.12	N/A <0.01	N/A <0.01	<0.17 0.04
trans-Nonachlordane	0.13	<0.02	0.12	<0.01	< 0.01	0.18
trans-Nonachiordane	0.11	<0.02 <0.02	0.09	<0.02 <0.04	<0.02 <0.05	
						0.04
cis-Chlordane Endrin	<0.01 <0.05	<0.02 <0.07	<0.02 <0.08	<0.01 0.02	<0.01 0.02	<0.01 <0.05
Endilli	\0.0 <i>5</i>	\0.07	₹0.00	0.02	0.02	\0.UJ
Lipid Smedes (%)	1.49	2.48	3.08	1.87	1.68	1.91

Notes: N/A: Samples not analysed

nd: Substances were not detected above the Limit of Detection (LOD) (Appendix 2)

< value: value = Limit of Quantitation (LOQ) for the relevant determinand

Appendix 1 (Page 3 of 4): Results of monitoring of shellfish-growing areas - 2002.

Sample site	Cromane, Castlemaine Harbour	Dungarvan	Glengarriff, Bantry Bay	Greencastle, Lough Foyle	Kilkieran	Killary Inner
M.I. Reference no.	ENV 02/065	ENV 02/085	ENV 02/063	ENV 02/072	ENV 02/060	ENV 02/057
Sampling date	28/08/02	10/10/02	27/08/02	01/10/02	20/08/02	20/08/02
Sampling time	12.00	10.20	15.30	16.30	9.30	11.00
Species sampled	M. edulis	C. gigas	M. edulis	M. edulis	O. edulis	M edulis
Number individuals	50	25	50	50	25	50
Method of cultivation	bed	trestle	rope	bed	bed	rope
Water Parameters						
Temperature (°C)	17.4	12.7	17.5	15.5	N/A	N/A
Salinity (psu)	27.3	30.3	31.0	30.4	N/A	N/A
pH	8.29	8.27	8.50	8.15	N/A	N/A
Suspended Solids (mg L ⁻¹)	3.10	123	5.80	6.50	4.40	8.10
Shellfish						
Shell length range (mm)	40.5-59.7	81.4-118	43.9-59.9	43.2-79.5	62.5-94.9	46.3-60.0
Shell mean length (mm)	51.7	104	51.3	51.0	72.3	52.5
Shell length std dev (mm)	3.86	9.07	3.65	5.82	6.63	3.53
Meat weight (%)	28.8	14.1	53.8	31.7	6.30	36.9
Shell weight (%)	71.2	85.9	46.2	68.3	93.7	63.1
Meat water content (%)	75.2	75.4	79.8	78.2	79.5	81.2
Metals mg kg ⁻¹ (ppm) wet v						
Cadmium	0.25	0.41	0.18	0.09	0.44	0.20
Chromium	0.25	0.20	< 0.19	0.29	< 0.19	< 0.19
Copper	2.18	9.03	1.07	2.38	12.1	1.53
Lead	0.12	0.25	0.07	0.16	0.24	0.11
Mercury Zinc	<0.03 24.8	nd 182	nd 16.7	<0.03 16.6	<0.03 348	<0.03 30.4
Nickel	0.38	0.15	0.18	0.39	<0.14	0.33
Silver	0.03	0.13	< 0.03	< 0.03	0.89	0.04
PCB's µg kg-1 (ppb) wet wt						
CB Congener 28	< 0.01	0.04	< 0.01	0.06	< 0.01	< 0.01
CB Congener 31	0.04	0.05	0.04	0.07	0.04	0.04
CB Congener 52	0.18	0.14	0.23	0.21	0.19	0.12
CB Congener 101	0.18	0.25	0.23	0.22	0.18	0.12
CB Congener 105	0.03	0.06	0.04	0.09	0.03	0.02
CB Congener 118	0.08	0.22	0.06	0.19	0.05	0.04
CB Congener 138	0.11	0.18	0.18	0.16	< 0.06	0.06
CB Congener 153	0.11	0.42	0.27	0.24	0.08	0.16
CB Congener 156	< 0.03	< 0.02	< 0.05	< 0.03	< 0.05	< 0.03
CB Congener 180	< 0.02	< 0.01	< 0.02	< 0.01	< 0.02	< 0.02
Organic pesticides µg kg ⁻¹	(ppb) wet wt.					
DDD - pp'	0.01	0.06	< 0.01	0.33	0.13	< 0.01
DDE - pp'	0.13	0.28	0.11	0.44	0.30	0.11
DDT - op'	0.03	< 0.02	0.06	< 0.02	0.04	0.03
DDT - pp'	0.18	< 0.12	0.14	< 0.14	0.14	0.16
Dieldrin	N/A	0.42	N/A	0.43	N/A	N/A
HCB	0.02	0.03	0.02	0.02	< 0.01	0.01
alpha-HCH	0.01	0.08	0.03	0.07	0.01	0.01
beta-HCH	N/A	< 0.19	N/A	<0.23	N/A	N/A
gamma-HCH	<0.01	0.04	<0.01	0.02	<0.01	<0.01
trans-Nonachlordane	<0.02	0.23	<0.03	0.12	<0.03	< 0.02
trans-Chlordane	<0.04	0.02	< 0.06	<0.02	< 0.06	< 0.05
cis-Chlordane Endrin	<0.01 <0.01	<0.02 <0.06	<0.01 <0.01	<0.02 <0.07	<0.01 0.05	<0.01 0.01
	.0.01	.0.00	\U.U1	\0.07	0.05	0.01

Notes: N/A:

Samples not analysed Substances were not detected above the Limit of Detection (LOD) (Appendix 2) nd:

< value: value = Limit of Quantitation (LOQ) for the relevant determinand

Appendix 1 (Page 4 of 4): Results of monitoring of shellfish-growing areas - 2002.

Sample site	Kilmakillogue, Kenmare River	Mulroy Bay	Quigley's Point, Lough Foyle	Roaring Water Bay	Tralee Bay, Inner	Wexford Harbour
M.I. Reference no.	ENV 02/066	ENV 02/070	ENV 02/069	ENV 02/062	ENV 02/068	ENV 02/084
Sampling date	28/08/02	02/10/02	01/10/02	27/08/02	28/08/02	9/10/02
Sampling time	9.30	9.30	16.30	12.00	16.30	11.00
Species sampled	M. edulis	M. edulis	O. edulis	M. edulis	O. edulis	M. edulis
Number individuals	50	50	25	50	25	50
Method of cultivation	rope	rope	bed	rope	bed	bed
Water Parameters						
Temperature (°C)	16.7	14.8	15.5	16.8	17.2	13.7
Salinity (psu)	31.8	30.7	30.4	33.6	31.9	30.2
pH	8.31	7.17	8.15	8.28	8.29	8.24
Suspended Solids (mg L ⁻¹)	3.50	16.5	6.50	3.70	7.70	108
Shellfish						
Shell length range (mm)	41.1-59.6	44.2-59.6	59.7-84.6	40.9-59.9	67.7-91.4	47.0-60.0
Shell mean length (mm)	52.2	51.4	73.9	54.9	77.3	54.5
Shell length std dev (mm)	4.86	2.97	5.49	3.74	6.15	3.68
Meat weight (%)	43.9	39.4	11.8	43.1	10.3	40.8
Shell weight (%)	56.1	60.6	88.2	56.9	89.7	59.2
Meat water content (%)	77.4	77.0	73.8	76.7	76.7	73.6
Metals mg kg ⁻¹ (ppm) wet w						
Cadmium	0.36	0.24	0.56	0.13	0.53	0.08
Chromium	< 0.19	< 0.19	< 0.19	0.29	0.22	0.25
Copper	1.68	1.36	27.9	1.34	20.8	1.24
Lead	0.11	< 0.06	0.14	0.18	0.09	0.34
Mercury	nd	nd	< 0.03	nd	< 0.03	< 0.03
Zinc	36.3	15.6	332	20.2	401	13.6
Nickel Silver	0.14 <0.03	<0.14 nd	<0.14 1.13	0.34 nd	<0.14 1.45	0.24 <0.03
PCB's µg kg ⁻¹ (ppb) wet wt.	₹0.03	IIG	1.13	na	1.43	\0.03
	.0.01	0.05	0.06	.0.01	-0.01	0.05
CB Congener 28	<0.01	0.05	0.06	< 0.01	< 0.01	0.05
CB Congener 31	0.03	0.06	0.06	0.05	0.03	0.07
CB Congener 52	0.10	0.15	0.15	0.12	0.13	0.16
CB Congener 101	0.16 0.03	0.12 0.05	0.26 0.08	0.14 0.02	0.15 0.03	0.27
CB Congener 119	0.03	0.03	0.08	0.02	0.03	0.08
CB Congener 118 CB Congener 138	0.08	< 0.10	0.24	0.09	0.11	0.23
CB Congener 158 CB Congener 153	0.14	<0.10	0.28	0.13	0.16	0.25 0.33
CB Congener 156	< 0.03	< 0.11	< 0.02	< 0.03	< 0.03	< 0.03
CB Congener 180	<0.03	< 0.03	< 0.02	< 0.03	< 0.03	< 0.03
Organic pesticides µg kg ⁻¹ (1						
DDD - pp'	0.03	0.02	0.51	< 0.01	0.06	0.18
DDE - pp'	0.21	0.21	0.65	0.02	0.36	0.68
DDT - op'	0.03	< 0.02	< 0.01	< 0.02	0.03	0.02
DDT - pp'	0.13	< 0.14	< 0.11	0.08	0.11	< 0.13
Dieldrin	N/A	0.33	0.51	N/A	N/A	0.19
НСВ	0.02	0.02	0.01	0.02	0.01	0.02
alpha-HCH	0.02	0.06	0.05	0.02	0.01	0.06
beta-HCH	N/A	< 0.22	< 0.17	N/A	N/A	< 0.21
датта-НСН	< 0.01	0.04	0.03	0.03	< 0.01	0.02
trans-Nonachlordane	0.02	0.02	0.17	< 0.02	< 0.02	< 0.02
trans-Chlordane	< 0.04	< 0.02	0.02	< 0.04	< 0.03	0.02
cis-Chlordane	< 0.01	< 0.02	< 0.01	< 0.01	< 0.01	< 0.02
Endrin	< 0.01	< 0.06	< 0.05	< 0.01	0.03	< 0.06
Lipid Smedes (%)	1.96	1.80	2.33	1.82	1.68	1.83

Notes: N/A: Samples not analysed

nd: Substances were not detected above the Limit of Detection (LOD) (Appendix 2)

< value: value = Limit of Quantitation (LOQ) for the relevant determinand

Appendix 2: Trace Metals - Limits of Detection (LOD) (mg kg⁻¹ wet weight).

Metal	LOD
Cadmium	0.004
Chromium	0.07
Copper	0.16
Lead	0.02
Mercury	0.01
Zinc	1.21
Nickel	0.06
Silver	0.01

Glossary and Abbreviations

Determinants

DDD 4,4'-dichlorodiphenyldichloroethane

DDE 1,1'-dichloro-2-(2-chlorophenyl)-2-(4-chlorophenyl)ethene

DDT 4,4'-dichlorodiphenyl-1,1,1-trichloroethane

HCBHexachlorobenzeneHCHHexachlorocyclohexaneOCPsOrganochlorine pesticidesPCBsPolychlorinated biphenyls

Cd Cadmium Cr Chromium Cu Copper Hg Mercury Pb Lead Z_n Zinc Ni Nickel Silver Ag

Species

M. edulisMytilis edulisBlue musselO. edulisOstrea edulisNative/flat oysterC. gigasCrassostrea gigasPacific oyster

Others

QUASH Quality assurance of sampling and sample handling (in marine environmental

monitoring)

QUASIMEME Quality assurance of information for marine environmental monitoring

RM Reference material

CRM Certified reference material

N/A
 LOD
 Limit of detection
 nd
 Not detected
 SI
 Statutory Instrument
 EU
 European Union

AA Atomic absorption (spectroscopy)

GC-ECD Gas chromatography with electron capture detection

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